THE CYTOGENETICS OF CAREX FLAVA AND ITS ALLIES*

By Elizabeth W. Davies

University College of Leicester

I. Introduction

II. Cytological Technique

III. Chromosome Numbers

IV. Hybrids and Hybridisation

A. Natural Hybrids

B. Artificial Hybrids

V. Discussion

VI. Conclusion and Summary

VII. Acknowledgments

1. INTRODUCTION

This group of closely allied species included in the section *Extensae* of the subgenus *Carex* is represented in Great Britain by : *Carex flava* L., *C. lepidocarpa* Tausch, *C. demissa* Hornem., *C. serotina* Mérat, and *C. scandinavica* E. W. Davies. The taxonomy of these species has caused difficulty for many years, and, although the chromosome numbers have been investigated by workers in Sweden, Japan and America, there has been considerable discrepancy in their results (see below), and no counts have previously been made on British material.

	Haploid		
Species	Chromosome Number	Determined by :	Date
C. extensa Good.	30	Wulff	1937
C. flava L.	29	Tanaka	1942, 1948
	30	Heilborn	1939
	30	Wahl	1940
C. lepidocarpa Tausch	29	Tanaka	1942, 1948
	34	Heilborn	1924
"C. oederi Retz" = C. serotina	Mérat 36	Heilborn	1922, 1924, 1928
	35	Tanaka	1948

2. Cytological Technique

The cytological technique employed throughout this investigation is a modification of the aceto-carmine squash method by the addition of a drop of iron alum. The *Carices* have exceedingly small chromosomes and hard wiry roots, which make them difficult cytological material.

The original attempts to count the chromosomes of this genus were made at mitosis using root tips. First embedding and sectioning methods were tried and later the root tip squash technique was used, staining with Feulgen, aceto-carmine, or a combination of the two stains together. However, the results throughout were unsatisfactory, as the cytoplasm was overstained and contained oil globules, while the chromosomes remained faint and aggregated together. Thus the counts were never accurate enough to detect a difference of two chromosomes at mitosis, which is essential when examining a genus

* Part of a thesis approved for the degree of Ph.D. by the University of London.

like *Carex* which has an aneuploid series with many allied species with consecutive chromosome numbers.

Finally young male spikes were fixed and meiosis was investigated using the modified aceto-carmine method. This produced well stained and spread chromosomes and, in contrast, a light background, as the cytoplasm absorbed little stain.

3. Chromosome Numbers

The chromosome numbers have perhaps not proved as valuable a criterion in the taxonomy of these species as had formerly been hoped. However, the chromosomes of the British members of the *Extensae* and of *C. mairii* Coss. & Germ., a closely allied species from southern Europe, have been examined from several localities, and are remarkably similar and uniform in number and morphology (Plates 12 and 13, figs. 1-14). This is further evidence that they are a naturally closely united group distinct from the other sections, such as *Distantes, Acutae, Montanae*, etc., which differ from them in size and morphology of their chromosomes.

The chromosome numbers of these species together with their subspecies and varieties are given in Table 1. The number of plants of each species and their localities is stated in each case.

Thus this aggregate of species forms a short aneuploid series with chromosome numbers ranging from 30 to 35. If this series is ascending, the low numbered members are assumed to be primitive, while the species with more numerous chromosomes are secondary and advanced. If this is the case, *C. flava* is the oldest and original member of this aggregate, and the other species are derived and more recent in origin. This aspect of the problem will be referred to, and elaborated, later, when the curious abnormalities, found at meiosis in hybrid sedges, have been discussed in the next section.

4. Hybrids and Hybridisation

The study of the interspecific hybrids of a critical group such as the C. flava aggregate is of great importance, as it frequently yields information about the relationships of the species, their age and evolution. The natural and artificial hybrids will now be discussed separately.

(a) Natural Hybrids

Natural hybrids are found between members of this aggregate, whenever two or more of the species grow together and their flowering periods overlap. However this is rather infrequent in this country, except in the case of the widespread C. demissa, and it appears that ecological barriers prevent much introgression between the species. The hybrids found in the field are listed in Table 2, and these will be discussed later, after some of the abnormalities seen at meiosis have been briefly outlined.

The meiosis of hybrid *Carices* is always highly irregular, and cytology produces conclusive evidence of hybridity in a suspected plant. However, the arrangement of the chromosomes and their behaviour at metaphase and anaphase is very different from what is usually seen in hybrids of most genera. First, and perhaps of most fundamental importance, there are no lagging chromosomes at anaphase I, but these can usually be seen to a small extent at anaphase II. Thus the normal sequence of events generally accepted for hybrids in most genera, with the possible exception of *Luzula*, is reversed. Secondly, at metaphase I, instead of the occurrence of tri- and bi-, or univalents, depending on the nature of the hybrids, there are frequently chains or even rings formed of from 3 to 6 or even 8 chromosomes (Plate 13, fig. 16) and round the periphery of the metaphase plate univalents are often seen.

		Haploid	Number of	
	Species	number	plants sampled	Locality
1.	C. extensa Good.	30	4	Rhos Neigr, Anglesey.
2.	C. flava L.	30	4	Roudsea Wood, Haverthwaite, Lancs.
		30	1	Malham Bog, Yorks.
		30	2	Preda, Graubünden, Switzerland.
		30	1	Lake Carezza, W. Dolomites, N. Italy.
3.	(a) C. lepidocarpa Tausch	34	1	Wicken Fen, Cambs.
	subsp. lepidocarpa	34	1	Fulbourn Pond, Cambs.
		34	3	Wittering Marsh, Northants.
		34	1	Gordale Scar, Malham, Yorks.
		34	1	Malham Tarn, Yorks.
		34	1	Moughton Scar, Yorks.
		34	3	Widdybank Fell, Teesdale, Durham.
		34	1	Gotland, Sweden.
		34	1	Tiefencastel, Switzerland.
3.	(b) C. lepidocarpa subsp. scotica	34	1	Cwm Glas, Snowdon, Caernarvon.
	E. W. Davies	34	1	Carn Eige, Cannich, Inverness.
		34	1	Inchnadamph, W. Sutherland.
		34	1	Preda, Graubünden, Switzerland.
		34	1	Albula, Graubünden, Switzerland.
4.	C. mairii Coss. & Germ.	34	4	Lumbreras, Spain.
5.	C. demissa Hornem.	35	2	Shapwick peat moor, Somerset.
		35	1	Bredon Cloud Wood, Leics.
		35	3	Strines Moor, Derbyshire.
		35	1	Little Dun Fen, Westmorland.
		35	3	Loch Tummel, Perth.
		35	2	Cannich, Inverness.
		35	1	Cwm Glas, Snowdon, Caern.
		35	1	Furnols, Echandelyse, Auvergne, France.
		35	1	Lac de la Faye, Auvergne, France.
		35	1	Issoire, Central France.
		35	1	Font Romeu, E. Pyrenees, France.
6.	C. serotina Mérat	35	1	L'Ancresse, Guernsey.
		35	1	Frensham Pond, Surrey.
		35	2	Fulbourn Pond, Cambs.
		35	2	Rhos Neigr, Anglesey.
		35	2	Hale Moss, Bentham, Lancs.
		35	1	Monea, Ireland.
		35	2	Hökatorp, Skåne, Sweden.
		35	2	Alvar, Öland, Sweden.
		35	2	Gotland, Sweden.
	C. serotina var. cyperoides	35	1	Shapwick peat moor, Somerset.
7.	C. scandinavica E. W. Davies	35	1	Sweden.
		35	1	Loch Torridon, W. Ross.

TABLE 1 Chromosome numbers of the C. flava aggregate

The meiosis appears to be more abnormal and irregular when the parents of the hybrid are very dissimilar, and have widely different chromosome numbers. The metaphase I, described above, represents a hybrid between C. demissa (n = 35) and C. hostiana (n = 28). In an interspecific hybrid between two more closely related species such as C. demissa (n = 35) and C. lepidocarpa (n = 34), or C. demissa (n = 35) and C. serotina (n = 35), meiosis is more normal, and apart from a few univalents and perhaps one conglomerate group of chromosomes forming a chain, the rest are arranged on the metaphase plate as bivalents (Plate 13, figs. 17-20). Thus a fairly high degree of homology

131

is indicated between these species with similar and consecutive chromosome numbers; for the pollen of hybrid plants is never completely sterile, and fertility usually ranges from 20 to 35 per cent (Table 2).

The hybrids detected in the field between British members of this aggregate will now be briefly discussed.

•			% Pollen	
			fertility	Localities
1. C. flava	×	C. lepidocarpa	29%	Lake Carezza, Dolomites, N. Italy.
2. C. flava	×	C. demissa	22%	Roudsea Wood, Haverthwaite, Lancs.
3a. C. lepidocarpa	×	C. demissa	22%	Teesdale, Durham.
subsp. <i>lepidocarpa</i>				
	Х	**	24%	Derby Fen, Norfolk.
**	×	,,	30%	S. Cerney, Glos.
,,	×	,,	35%	Hökatorp, Skåne, Sweden.
,,	Х	,,	25%	Krogenbund Mose, Zealand, Denmark.
b. C. lepidocarpa	Х	C. demissa	32%	Loch Tummel, Perth.
subsp. scotica				
,,	×	,,	35%	Creag-na-Caillich, Perth.
,,	×	**	33%	Creag-an-Lochan, Perth.
,,	×		35%	Ben-y-Vrackie, Pitlochry, Perth.
,,	×	**	28%	Ben Dorain, Argyll.
**	×	**	30%	Glen Doll, Clova, Angus.
**	×	**	31%	Cwm Glas, Snowdon, Caernarvon.
4. C. lepidocarpa	×	C. serotina	25%	Wicken fen, Cambs.
,,	×	,,	20%	S. Cerney, Glos.
5a. C. demissa	×	C. serotina	29%	S. Cerney, Glos.
b. C. demissa	X	C. serotina var. cyperoides	26%	Shapwick, Somerset.
6a. C hostiana	X	C. lepidocarpa	0%	Chippenham Fen, Cambs.
		subsp. lepidocarpa		
b. ,,	X	C. lepidocarpa	0%	Loch Tummel, Perth.
		subsp. scotica		
. ,,,	×	**	0%	Waulk Mill Bay, Mainland, Orkney.
				Lech, Voralberg, Austria.
7. C. hostiana	×	C. demissa	0%	Loch Tummel, Perth.
8. C. distans	×	C. lepidocarpa	0%	Totternhoe, Beds.
-		_		

TABLE 2 Natural hybrids in the C. flava aggregate

(1) C. flava

This rare British plant, with its specialised ecological requirements, seldom meets the other members of the group in this country. However, at Roudsea Wood (Lancashire), C. demissa occurs with C. flava, and there is a range of intermediate forms, which are clearly of hybrid origin. These plants revealed a highly irregular meiosis, and the pollen showed between 70 and 80 per cent sterility. It certainly seemed as if back crosses with the parent species were present as well as F_1 plants, for there was a great range of forms and it appeared that, in time, C. demissa would completely oust C. flava by hybridisation, for the latter species was scarce compared with the abundance of hybrid plants and C. demissa. This is the only record of a C. flava hybrid from this country; as this species has never been found growing with or near C. lepidocarpa, C. serotina or C. scandinavica, and as their ecological requirements are rather different, hybrids between C. flava and these species seem unlikely to occur naturally. However, hybrids and back crosses between this species and C. lepidocarpa were abundant by Lake Carezza in the Western Dolomites. Here, in North Italy, the habitat and the topography seemed suitable for both species and they were hybridising freely.

(2) C. lepidocarpa

Subsp. lepidocarpa frequently hybridises with C. demissa, and a hybrid swarm often predominates, while the parent species are comparatively rare, and sometimes difficult to detect. This certainly is the case on the lower slopes of Widdybank Fell in Upper Teesdale (Plate 13, fig. 17) and at Hökatorp (Plate 13, fig. 18), a calcareous fen by the sea in S.E. Skåne, Sweden. Likewise, on base-rich Scottish and Welsh mountains hybrid swarms between C. demissa and subsp. scotica are common at about 1,000 m. where the distribution areas of these two species overlap, whereas below and above this altitude the two species remain quite distinct.

In the same way hybrids might be expected between C. lepidocarpa and C. serotina in the few calcareous localities where these species are known to occur together. However, this hybrid is rare, apparently because C. serotina flowers and fruits at least one month later than the other four species. At Fulbourn Pond (Cambridgeshire), where C. lepidocarpa and C. serotina grow completely intermixed, there is no sign of hybridisation, and, in late June, the former species is in ripe fruit while the latter is still in flower.

Consequently, it was rather unexpected to find a plant at Wicken Fen, Cambridgeshire, which appeared to be a hybrid of this parentage, growing with C. serotina and C. lepidocarpa. This plant had an irregular meiosis and the pollen showed 80 per cent sterility. The only other locality examined where these two species were found growing together is a disused gravel pit on the Cotswolds near S. Cerney, and, as C. demissa also grows at the edge of the pit, the situation is complicated further. A complex hybrid swarm involving all three species occurs, and the range of forms is very great. Although the majority of plants seem to be intermediate between C. lepidocarpa and C. serotina, and some forms contain C. demissa also, there is a large stand of pure C. serotina at one side of the gravel pit. This can almost certainly be correlated with the late flowering period of this species, for specimens of pure C. lepidocarpa and C. demissa were not seen and these species had clearly been ousted by hybridisation.

(3) C. demissa

This species, which is tolerant of a wide range of habitats, is frequently found growing with, or near, the other species, and hybrids are fairly common.

The hybrids with C. flava and C. lepidocarpa have already been mentioned, but on Shapwick peat moor in Somerset, this species forms a hybrid swarm with C. serotina var. cyperoides. The latter is rather uncommon, and it seems likely that it may, in time, be lost completely, if hybridisation with C. demissa continues, for an enormous range of intermediate forms is now abundant on the moor, and C. serotina var. cyperoides is becoming increasingly rare. The cytology of this hybrid has been examined, and reveals a highly irregular meiosis (Plate 13, fig. 19), with a conglomerate ring of chromosomes surrounded by univalents.

(4) C. serotina

The few hybrids that are known between C. serotina and the other members of the aggregate have already been discussed, and it undoubtedly seems to be the late flowering of this species that acts as the main barrier, and keeps it distinct from the other members of the group.

(5) C. scandinavica

C. scandinavica, in the British Isles confined to northern and western Scotland, has

133

as yet never been found growing with any of the other species in this country. Hence no hybrids have been recorded.

(6) C. hostiana and C. distans

This study would not be complete without mentioning C. hostiana DC. and C. distans L., two members of the closely related section Distantes, for the former species very frequently hybridises with C. lepidocarpa and C. demissa. C. hostiana (n = 28), like C. demissa, grows in a wide range of habitats and tolerates many different soil conditions, which probably accounts for the frequency of its hybrids.

The hybrid between C. hostiana and C. lepidocarpa is sufficiently well known to have a name (C. × xanthocarpa), and is of fairly frequent occurrence. It grows (Table 2) at Chippenham Fen, Cambridgeshire, where the hybrid forms a large clone, which appears to multiply entirely vegetatively and now covers a considerable area, surrounded by the parent species.

Likewise C. hostiana hybridises with C. demissa. This hybrid was collected by Loch Tummel, Perthshire, and the plant showed a highly irregular meiosis with several chains of chromosomes and numerous univalents.

Lastly a hybrid between C. distans (n = 37) (Plate 13, fig. 15) and C. lepidocarpa was recorded from Cow Common, Totternhoe, Bedfordshire. This hybrid formed a clonal stand like $C. \times$ xanthocarpa, and had highly sterile pollen and a very irregular meiosis.

Thus it seems that all the members of this aggregate are capable of hybridisation, and considerable gene-flow between species is possible. It seems also that it is not genetical incompatibility but ecological and topographic barriers that keep C. flava, C. lepidocarpa, C. demissa and C. scandinavica apart, as distinct entities, while, in contrast, late seasonal periodicity seems to play the major role in the case of C. serotina.

(b) Artificial Hybrids

These flavoid species of *Carex*, like all those in the subgenus *Carex*, lend themselves to artificial hybridisation, as the male spike is easily removed without damaging the female organs. The crosses were carried out during May and early June, and the ripe seeds collected at the end of July and sown immediately.

The artificial hybridisations included all possible combinations between the five British members of the aggregate, and some crosses between these and C. mairii, C. extensa and C. hostiana (Table 3). However, although in most combinations some apparently ripe seed was produced, germination is slow and spasmodic in this genus, so as yet only a few of the hybrid seedlings have produced fertile spikes. These are listed in Table 3, and Plate 13, fig. 20 shows the very irregular meiosis of these artificial hybrids.

Thus it would seem that these five British species included in the C. flava aggregate, are capable of considerable gene-flow from one to another, and produce F_1 plants with between 20 and 30 per cent good pollen. They should therefore be regarded as ecospecies, and the group as a whole as one coenospecies (Gregor, 1939; Clausen, Keck & Hiesey, 1939).

6. DISCUSSION

The cytology, evolution and origin of the aneuploid series in the genus *Carex* will be discussed more fully in another paper.

However, it seems reasonable to postulate from the present cytological, ecological (Davies, 1954) and taxonomic (Davies, 1953a, b, c) evidence that, within this coenospecies,

	For als Derent		Mala Davant	No. of	%	Pollen Fertility
-	remaie Parent		Male Parent	seeanngs		oj nyona
1.	C. jiava	Х	Leal Transa	4		20
~	Roudsea, Lancs.	. /	Loch Tummel, Pertn.	4		20
2.	C. flava	Х	lepiaocarpa subsp. scotica	-		
~	Roudsea, Lancs.		Cannich, Inverness.	1		
3.	C. lepidocarpa	Х	flava			
	Tiefencastel, Switzerland.		Preda, Switzerland.	4		
4.	C. demissa	Х	flava			
	a. France		Roudsea, Lancs.	1		<u> </u>
	b. Bredon, Leics.		Roudsea, Lancs.	2		_
5.	C, demissa	х	lepidocarpa			
	a. Shapwick, Somerset.		Teesdale, Durham.	5		25
	b. France		Wicken, Cambs.	3		
	c. Wild Boar Fell, Westmorland		Wittering, Northants.	5		. —
6.	C. demissa	Х	C. serotina			
	a. Strines Moor, Derbyshire.		Anglesey.	2		29
	b. France.		Hökatorp, Sweden.	1		30
	c. Shapwick, Somerset.		Anglesey.	5		
	d. France.		Monea, Ireland.	1		
7.	C. demissa	Х	scandinavica	3		\rightarrow
	Strines Moor, Derbyshire.		Sweden.			
8.	C. demissa	х	mairii			
	Bredon, Leics.		Lumbreras, Spain.	2		
9.	C. serotina	Х	lepidocarpa	6		24
	Hökatorp, Sweden.		Pyrenees.			
10.	C. mairii	×	lepidocarpa subsp. lepidocarpa	1		_
	a. Pyrenees.		Gordale, Yorks.			
	C. mairii	х	lepidocarpa subsp. scotica	2		<u> </u>
	b. Pyrenees.		Cannich, Inverness.			
11.	C. mairii	х	serotina			
	Pyrenees.		Fulbourn, Cambs.	2		
12.	C. extensa	x	serotina			
	Anglesev.		Sweden.	1		
13.	C. hostiana	×	lepidocarpa	-		
	Teesdale.		Pyrenees.	2		·
	2000000			-		

TABLE 3 Artificial hybrids in *C. flava* aggregate

C. flava is the relict and primitive member, with the lowest chromosome number, specialised ecological requirements and relict distribution (Davies, 1953 d). The development of the other ecospecies from C. flava has been accompanied by an increase in chromosome numbers, a greater variation within the species and an ability to grow in a wider range of plant communities. Thus C. lepidocarpa, with four more chromosomes, is more abundant, adaptable and probably of more recent origin, although not so recent as the very widespread and variable C. demissa with a capacity to flourish under various ecological conditions. Lastly, C. serotina, with the same chromosome number as C. demissa, yet requiring more specialised conditions, seems to be a more primitive and possibly a reduced form; which needs the low sparse vegetation of an open habitat with little competition.

It seems likely that this species, with the widest geographical distribution (Davies, 1953 d) of the group, spread rapidly during and after the last glaciation, recolonising the open habitats produced by the retreating ice. Within recent times, owing to its specialised, requirements, this species has become confined to scattered and local areas; probably the presence of different forms which are not true ecotypes (as these differences do not seem

135

ELIZABETH W. DAVIES

related to ecology) can be correlated with this fact. The isolated populations throughout the British Isles show minor differences, which are retained in cultivation; the most striking examples of such isolated populations are some of the forms which are found in Ireland and the Baltic Islands of Gotland and Öland, for these plants are almost worthy of recognition as distinct taxa. This continued isolation is probably the reason for the existence of a form on Shapwick peat moor, Somerset, sufficiently different to be considered a variety (*C. serotina* var. *cyperoides*). Further results of this isolation are possibly to be seen in the origin of *C. scandinavica* and *C. viridula* Michx. (Davies 1953 d). These two plants, which are rather similar to *C. serotina* but have different and more limited geographical distributions, have only recently been considered distinct species. They may have been evolved within comparatively recent times owing to geographic separation.

6. Conclusion and Summary

A cytogenetical survey of the C. flava aggregate (C. flava L., C. lepidocarpa Tausch, C. demissa Hornem., C. serotina Mérat, C. scandinavica E. W. Davies) is made, and the origin and evolution of the species discussed.

These five members of section *Extensae* and *C. mairii* are found to form a short aneuploid series, with their haploid numbers ranging from n = 30 to n = 35.

The interspecific hybrids between the members of this group, with the exception of C. demissa, were found to be rather uncommon. This seems to be due to the different and specialised ecological requirements of the species, topographic separation and the late flowering season of C. servina. Hybrids were examined and synthesised whenever possible, and in every case the pollen showed at least 20 per cent fertility; this increased when the parent species had the same or nearly similar chromosome numbers.

Thus these five closely related species, which are capable of considerable gene-flow from one to another, and produce natural and artificial hybrids with between 20 and 30 per cent good pollen, are regarded as ecospecies and the group as a whole as one coenospecies.

C. flava seems to be a relict and stable plant with an isolated and scattered distribution. C. lepidocarpa, with its three subspecies, namely subsp. lepidocarpa and subsp. scotica, which form a distconinuous topocline in the British Isles, and subsp. jemtlandica, so far only known from Scandinavia, is more widespread.

The abundant C. demissa appears to be a variable plant, but the range of variation is continuous, and within the limits of one adaptable species. In contrast, C. serotina, with its local and fragmentary distribution, has a number of forms which, owing to long isolation, seem to give rise to varieties, such as var. cyperoides, and eventually species; such is probably the origin of C. scandinavica and C. viridula.

7. Acknowledgments

I have pleasure in recording my gratitude to Professor T. G. Tutin for his advice and encouragement throughout this work. I should also like to express my thanks to the Research Board, University College, Leicester, for the award of a scholarship during the period 1950-53, and for giving considerable financial help towards my field studies during this period.

REFERENCES

ANDERSON, E., 1949, Introgressive hybridization. New York.

BAKER, H. G., 1948, Stages in invasion and replacement demonstrated by species of Melandrium, J. Ecol., 36, 96-110.

-, 1951, Hybridization and natural gene-flow between higher plants, Biol. Rev., 26.

CLAUSEN, J., KECK, D. D. & HIESEY, W. M., 1939, The concept of species based on experiment, Amer. J. Bot., 26, 103-106.

PLATE 12

Figure 1 shows a pollen grain mitosis and two of the three aborting nuclei, and Figs. 2-9 show meiosis in the pollen mother cells. All photographs \times 2,400.



- Fig. 2. C. flava, Malham Tarn Moss, Yorks., n = 30.
- Fig. 3. C. flava, Preda, Graubünden, Switzerland, n = 30.
- Fig. 4. C. lepidocarpa subsp. lepidocarpa, Wicken Fen, Cambs., n = 34.
- Fig. 5. C. lepidocarpa subsp. scotica, Inchnadamph, Sutherland, n = 34.
- Fig. 6. C. demissa, Loch Tummel, Perthshire, n = 35.
- Fig. 7. C. demissa, Furnols, Central France, n = 35.
- Fig. 8. C. serotina, calcareous marsh, Fulbourn, Cambs., n = 35.
- Fig. 9. C. serotina, salt marsh, Hökatorp, Sweden, n = 35.

PLATE 13

Figures 10-15 show normal meiosis in the pollen mother cells, and Figs. 16-20 show highly irregular meiosis in the pollen mother cells of some hybrid sedges, with univalents marked by arrows, and a few multivalents forming chains and rings of chromosomes. All photographs \times 2,400.



- Fig. 13. C. mairii, Lumbreras, Spain, n = 34.
- Fig. 14. C. extensa, Rhos Neigr, Anglesey, n = 30.
- Fig. 15. C. distans, St. Nectaire, Central France, n = 37.
- Fig. 16. C. demissa × C. hostiana, Loch Tummel, Perthshire.
- Fig. 17. C. lepidocarpa × C. demissa, Teesdale, Durham.
- Fig. 18. C. lepidocarpa × C. demissa, Hökatorp, Sweden.
- Fig. 19. C. demissa × C. serotina var. cyperoides, Shapwick, Somerset.
- Fig. 20. C. demissa, Shapwick, Somerset, × C. lepidocarpa, Teesdale, Durham; hybridised 10 May, 1951.

DAVIES, E. W., 1953a, Notes on Carex flava and its allies : I. A sedge new to the British Isles, Watsonia, 3, 66-69.

------, 1953b, Notes on Carex flava and its allies : II. Carex lepidocarpa in the British Isles, Watsonia, 3, 70-73.

————, 1953c, Notes on Carex flava and its allies : III. The taxonomy and morphology of the British representatives, *Watsonia*, **3**, 74-79.

- ------, 1953d, Notes on Carex flava and its allies : IV. Geographic distribution, Watsonia, 3 80-84.
- GREGOR, J. W., 1942, The units of experimental taxonomy, Chronica Botanica, 7, 193.
 - -----, 1944, The ecotype, Biol. Rev., 19.
 - -----, 1946, Ecotypic differentiation, New Phytol., 45, 254.
- HUXLEY, J. S. (ed.), 1940, The New Systematics. Oxford.
- KÜKENTHAL, G., 1900, Cyperaceae-Caricoideae, Das Pflanzenreich, IV, 20.
- LÖVE, Á. & D., 1948, Chromosome numbers of Northern Plant Species. Reykjavik.
- NELMES, E., 1947, Two critical groups of British Sedges, Rep. Bot. Soc. & E.C., 13, 95-99.
- ———, 1949, Carex flava and its allies, in Wilmott, A. J. (ed.), British Flowering Plants and Mo dern Systematic Methods, 85. London.
- SENAY, P. 1950, Le groupe des Carex flava et C. oederi, Bull. Mus. Nat. Hist. Nat., Paris, 2me série, 22, 618-624, 790-796.
 - -----, 1951, Le groupe des Carex flava et C. oederi, Bull. Mus. Nat. Hist. Nat., Paris, 2me série, 23, 146-152.
- STEBBINS, G. L., 1950, Variation and Evolution in Plants. Oxford.
- TISCHLER, G., 1950, Die Chromosomenzahlen der Gefässpflanzen Mitteleuropas.
- TURRILL, W. B., 1946, The ecotype concept, a consideration with appreciation and criticism, especially of recent trends, New Phytol., 45, 34.
- TUTIN, T. G., 1952, Cyperaceae, in Clapham, A. R., Tutin, T. G. & Warburg, E. F., Flora of the British Isles. Cambridge.
- VALENTINE, D. H., 1948, Studies in British Primulas : II. Ecology and taxonomy of primrose and oxlip (Primula vulgaris Huds. and Primula elatior Schreb.), New Phytol., 47, 111.

-, 1949, The units of experimental taxonomy, Acta Biotheoretica, 9, 75.

WAHL, H. A., 1940, Chromosome numbers and meiosis in the genus Carex, Amer. J. Bot., 27, 458.